

AQUATIC ANIMAL DISEASE REPORT - 2021

Country/territory: PHILIPPINES		Disease status/occurrence code a/b/												Level of diagnosis	Epidemiological comment numbers	
Item	Month															
DISEASES PREVALENT IN THE REGION																
FINFISH DISEASES																
OIE-listed diseases																
1. Infection with epizootic haematopoietic necrosis virus	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000				
2. Infection with infectious haematopoietic necrosis virus	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000				
3. Infection with spring viraemia of carp virus	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000				
4. Infection with viral haemorrhagic septicaemia virus	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000				
5. Infection with <i>Aphanomyces imadans</i> (EUS)																
6. Infection with red sea bream iridovirus	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000			I	1
7. Infection with koi herpesvirus	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000			I, III	2
Non OIE-listed diseases																
8. Grouper iridoviral disease	-	-	-	-	-	-	-	-	-	-	-	-				
9. Viral encephalopathy and retinopathy	-	-	-	-	-	-	-	-	-	-	-	-			I, III	3
10. Enteric septicemia of catfish	***	***	***	***	***	***	***	***	***	***	***	***				
11. Carp Edema Virus Disease	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000				
12. Tilapia lake virus (TLV)	+	+	-	+	+	+	-								I, III	4
MOLLUSC DISEASES																
OIE-listed diseases																
1. Infection with <i>Bonamia exitiosa</i>	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000				
2. Infection with <i>Perkinsus olseni</i>	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000				
3. Infection with abalone herpesvirus	***	***	***	***	***	***	***	***	***	***	***	***				
4. Infection with <i>Xenohaliotis californiensis</i>	***	***	***	***	***	***	***	***	***	***	***	***				
5. Infection with <i>Bonamia ostreae</i>	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000				
Non OIE-listed diseases																
6. Infection with <i>Marteilioides chungmuensis</i>	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000				
7. Acute viral necrosis (in scallops)	***	***	***	***	***	***	***	***	***	***	***	***				
CRUSTACEAN DISEASES																
OIE-listed diseases																
1. Infection with Taura syndrome virus	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000			I, III	5
2. Infection with white spot syndrome virus	+	+	+	+	+	+	+	+	+	+	+	+			I, III	6
3. Infection with yellow head virus genotype 1	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000			I, III	7
4. Infection with infectious hypodermal and haematopoietic necrosis virus	+	+	+	+	+	+	+	+	+	+	+	+			I, III	8
5. Infection with infectious myonecrosis virus	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000			I, III	9
6. Infection with <i>Macrobrachium rosenbergii</i> nodavirus (White Tail disease)	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000			I, III	10
7. Infection with <i>Hepatobacter penaei</i> (Necrotising hepatopancreatitis)	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000			I, III	10
8. Acute hepatopancreatic necrosis disease (AHPND)	+	+	+	+	+	+	+	+	+	+	+	+			I, III	11
9. Infection with <i>Aphanomyces astaci</i> (Crayfish plague)	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000				
Non OIE-listed diseases																
10. Hepatopancreatic Microsporidiosis caused by <i>Enterocytozoon hepatopenaei</i> (HPM-EHP)	+	+	+	+	+	+	+	+	+	+	+	+			I, III	12
11. Viral covert mortality disease (VCMD) of shrimps	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000				
12. <i>Sirrolasma eriocheiris</i> infection	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000				
13. Decapod iridescent virus 1 (DIV-1)	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000	0000				
AMPHIBIAN DISEASES																
OIE-listed diseases																
1. Infection with <i>Ranavirus</i> species	***	***	***	***	***	***	***	***	***	***	***	***				
2. Infection with <i>Batrachochytrium dendrobatidis</i>	***	***	***	***	***	***	***	***	***	***	***	***				
3. Infection with <i>Batrachochytrium salamandrivorans</i>	***	***	***	***	***	***	***	***	***	***	***	***				

Prepared by:

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Position: Veterinaria IV

Date: June 14, 2021

ANY OTHER DISEASES OF IMPORTANCE												
1												
2												

DISEASES PRESUMED EXOTIC TO THE REGION⁸ LISTED BY THE OIE
Fishfish: Infection with HPR-deleted or HPR0 salmon anaemia virus; Infection with salmon pancreas disease virus; Infection with *Gyrodactylus salaris*.
Molluscs: Infection with *Marteilia refringens*; *Perkinsus marinus*.

NOT LISTED BY THE OIE
Fishfish: Channel catfish virus disease

a) Please use the following occurrence code:

Occurrence code and symbol	Definition	Occurrence code and symbol	Definition
Disease present =	The disease is present with clinical signs in the whole country (in domestic species or wildlife)	Disease absent -	The disease was absent in the country during the reporting period (in domestic species or wildlife).
Disease limited to one or more zones =()	The disease is present with clinical signs, and limited to one or more zones/compartments (in domestic species or wildlife)	Never reported 0000	The disease has "never been reported" (historically absent) for the whole country in domestic species and wildlife.
Infection/infestation +?	Confirmed infestation or infection using diagnostic tests, but no clinical signs observed (in domestic species or wildlife)	No information ***	No information is available regarding the presence or the absence of this disease during the reporting period (in domestic species or wildlife).
Infection/infestation limited to one or more zones =?()	Confirmed infestation or infection using diagnostic tests, but no clinical signs observed and limited to one or more zones/compartments (in domestic species or wildlife)		
Disease suspected ?	The presence of the disease was suspected but not confirmed (in domestic species or wildlife)		
Disease suspected but not confirmed and limited to one or more zones =?()	The presence of the disease was suspected but not confirmed and limited to one or more zones/compartments (in domestic species or wildlife)		

b) If there is any changes on historical data, please highlight in RED.

1. Epidemiological comments:
(Comments should include: 1) Origin of the disease or pathogen (history of the disease); 2) Species affected; 3) Disease characteristics (usual clinical signs or lesions); 4) Pathogen (isolated/sero-typed); 5) Mortality rate (high/low; decreasing/increasing); 6) Death toll (economic loss, etc.); 7) Size of infected areas or names of infected areas; 8) Preventive/control measures taken; 9) Samples sent to national or international laboratories for confirmation (indicate the names of laboratories); 10) Published paper (articles in journals/website, etc.) and 11) Unknown diseases: describe details as much as possible.)

Comment No.	
1	<p>Infection with <i>Aphanomyces invadans</i> (EUS) First Quarter: EUS was not detected by gross morphological examination in <i>Anguilla</i> spp. (elver), <i>A. bicolor</i> (elver) and <i>A. bicolor pacificus</i> (juvenile) from Nueva Ecija, Batangas and Tarlac. Examinations were conducted by BFAR Central and Regional Laboratories. Second Quarter: EUS was not detected by gross morphological examination in <i>Anguilla</i> spp. (grow-out) from Batangas and Cabadbaran. Examination was conducted by BFAR Central Fish Health Laboratory.</p>
2	<p>Red Sea Bream Iridoviral Disease (RSID) First Quarter: Tilapia (fry) and Pompano (grow-out) analyzed using PCR test showed negative results for Red Sea Bream Iridoviral Disease. Samples were collected from Iloilo and Nueva Ecija. Examinations were conducted by BFAR Central, Regional and Southeast Asian Fisheries Development Center (SEAFDEC) Fish Health Laboratories. Second Quarter: Milkfish (fingerling and grow-out) and Tilapia (fingerling and grow-out) analyzed using PCR test showed negative results for Red Sea Bream Iridoviral disease. Samples were collected from Iloilo, Davao del Sur, Davao City, Nueva Ecija and Batangas. Examination was conducted by SEAFDEC and BFAR Central Fish Health Laboratory.</p>
3	<p>Viral Encephalopathy and Retinopathy (VER) First Quarter: Pompano (fingerlings, grow-out), tilapia (fry) analyzed using PCR showed negative results for Viral Encephalopathy and Retinopathy. Samples were collected from Iloilo and Nueva Ecija. Examinations were conducted by BFAR Central and (SEAFDEC) Fish Health Laboratory. Second Quarter: Milkfish (fingerling and grow-out), <i>Siganus guttatus</i>, and Tilapia (fingerling and grow-out) analyzed using PCR tests showed negative results for Viral Encephalopathy and Retinopathy. Samples were collected from Iloilo, Davao City, Nueva Ecija and Batangas. Examinations were conducted by SEAFDEC and BFAR Central Fish Health Laboratories.</p>
4	<p>Tilapia Lake Virus (TILV) First Quarter: Origin of the disease or pathogen (history of the disease) - detected 3 farms Species affected: Tilapia (fingerlings, fry, juvenile) Pathogen: Tilapia Lake Virus Size of infected areas or names of infected areas: La Union, Nueva Ecija and Agusan del Norte Samples sent to national or international laboratories for confirmation (indicate the name of the laboratories): Polymerase Chain Reaction (PCR) Test / BFAR Regional Fish Health Laboratory Second Quarter: Origin of the disease or pathogen (history of the disease) - detected in 2 farms Species affected: Tilapia (fingerlings, grow-out and broodstock) Pathogen: Tilapia Lake Virus Size of infected areas or names of infected areas: Bohol and Laguna Samples sent to national or international laboratories for confirmation (indicate the name of the laboratories): Polymerase Chain Reaction Test (PCR) / BFAR Central and BFAR Regional Fish Health Laboratory</p>
5	<p>Taura Syndrome (TS) First Quarter: <i>P. vannamei</i> (broodstock, adult, grow-out, juvenile, post-larvae, fry and nauplii) analyzed using PCR test showed negative for Taura Syndrome. Samples were collected from Butuan, Iloilo, Bulacan, Leyte, Oriental Mindoro, Cagayan, Pangasinan, Zambales, Quezon, Sorsogon, Camarines Norte, Bohol and Cebu. Examinations were conducted by BFAR Central and Regional Fish Health Laboratories. Second Quarter: <i>P. monodon</i> (grow-out and post-larvae) and <i>P. vannamei</i> (fry, nursery, post-larvae, adult, grow-out and broodstock) analyzed using PCR tests showed negative for Taura Syndrome. Samples were collected from Bulacan, Cagayan, Cebu, Batangas, Davao del Sur, Oriental Mindoro, Zambales, Bohol, Pangasinan, Nueva Ecija, Quezon, and Iloilo. Examinations were conducted by BFAR Central and BFAR Regional Fish Health Laboratories.</p>
6	<p>White Spot Disease (WSD) First Quarter: Origin of the disease or pathogen (history of the disease) - detected 3 farms Species affected: <i>P. monodon</i> (broodstock and post-larvae) and <i>P. vannamei</i> (grow-out and juvenile) Pathogen: White Spot Syndrome Virus Size of infected areas or names of infected areas: Negros Occidental, Leyte, Cagayan, Mindoro, Sorsogon, Camarines Norte, Davao del Sur, Agusan del Norte, Misamis Oriental, Surigao del Sur Samples sent to national or international laboratories for confirmation (indicate the name of the laboratories): Polymerase Chain Reaction (PCR) Test / BFAR Central, Regional and SEAFDEC Fish Health Laboratories. Second Quarter: Origin of the disease or pathogen (history of the disease) - detected in 17 farms Species affected: <i>P. Vannamei</i> (post-larvae, grow-out and spawner), <i>P. monodon</i> (post-larvae, grow-out and spawner), <i>S. olivacea</i> (grow-out) and <i>P. indicus</i> (grow-out) Pathogen: White Spot Syndrome Virus Size of infected areas or names of infected areas: Iloilo, Negros Occidental, Oriental Mindoro, Cagayan, Agusan del Norte, Sorsogon, Camarines Norte, Cebu, Leyte and Leyte Samples sent to national or international laboratories for confirmation (indicate the name of the laboratories): Polymerase Chain Reaction (PCR), Conventional, and IQ Plus tests/ BFAR Central, BFAR Regional and SEAFDEC Fish Health Laboratories.</p>
7	<p>Infection with Yellow Head Virus Genotype 1 (YHV) First Quarter: <i>P. monodon</i> (fry and grow-out) and <i>P. vannamei</i> (broodstock and grow-out) analyzed using PCR test showed negative results for Yellow Head Virus Genotype 1. Samples were collected from Butuan, Iloilo, Bulacan, Leyte and Mindoro. Examinations were conducted by BFAR Central and Regional Fish Health Laboratories. Second Quarter: <i>P. monodon</i> (post-larvae, grow-out and broodstock) and <i>P. vannamei</i> (post-larvae, grow-out, adult and broodstock) analyzed using Conventional test showed negative result for Yellow Head Virus. Samples were collected from Bulacan, Cagayan, Cebu, Batangas, Davao del Sur, Oriental Mindoro, Zambales, Pangasinan, Bohol, Iloilo and Misamis Occidental. Examination was conducted by BFAR Central Fish and BFAR Regional Health Laboratories.</p>
8	<p>Infection with Infectious Hypodermal and Haematopoietic Necrosis Virus (IHHNV) First Quarter: Origin of the disease or pathogen (history of the disease) - detected 6 farms Species affected: <i>P. monodon</i> (post larvae) and <i>P. vannamei</i> (post-larvae, adult) Pathogen: Infectious Hypodermal and Haematopoietic Necrosis Virus Size of infected areas or names of infected areas: Pampanga, Zambales, Quezon, Camarines Norte, Bohol and Cebu Samples sent to national or international laboratories for confirmation (indicate the name of the laboratories): Polymerase Chain Reaction (PCR) Test / BFAR Regional Fish Health Laboratories. Second Quarter: Origin of the disease or pathogen (history of the disease) - detected in 9 farms Species affected: <i>P. monodon</i> (post-larvae and grow-out) and <i>P. vannamei</i> (fry and adult) Pathogen: Infectious Hypodermal and Haematopoietic Virus Size of infected areas or names of infected areas: Bulacan, Zambales, Sorsogon, and Camarines Norte Samples sent to national or international laboratories for confirmation (indicate the name of the laboratories): Polymerase Chain Reaction (PCR) / BFAR Central and BFAR Regional Fish Health Laboratories.</p>
9	<p>Infectious Myonecrosis (IMN) First Quarter: <i>P. monodon</i> (fry, grow-out) and <i>P. vannamei</i> (broodstock, grow-out, adult) analyzed using PCR test showed negative for Infectious Myonecrosis. Samples were collected from Butuan, Iloilo, Bulacan, Leyte, Mindoro, Cagayan, Bohol and Cebu. Examinations were conducted by BFAR Regional Fish Health Laboratory. Second Quarter: <i>P. monodon</i> (post-larvae, grow-out and adult) and <i>P. vannamei</i> (nursery, post-larvae, grow-out, broodstock and breeders) analyzed using PCR tests showed negative for Infectious Myonecrosis. Samples were collected from Bulacan, Cagayan, Cebu, Batangas, Davao del Sur, Oriental Mindoro, Pangasinan, Zambales, Quezon, Iloilo and Bohol. Examinations were conducted by BFAR Central and BFAR Regional Fish Health Laboratories.</p>

10	<p>Neurotising Hepatopancreatitis (NHP) First Quarter: <i>P.monodon</i> (post-larvae, grow-out) and <i>P.vannamiei</i> (broodstock, grow-out, post-larvae) analyzed using PCR test showed negative for Neurotising Hepatopancreatitis. Samples were collected from Butuan, Boilo, Bulacan, Leyte, Mindoro, Quezon, Bohol and Cebu. Examinations were conducted by BFAR Central Fish Health Laboratory.</p> <p>Second Quarter: <i>P. vannamiei</i> (post-larvae, breeders and broodstock) and <i>P. monodon</i> (grow-out) analyzed using PCR tests showed negative for Neurotising Hepatopancreatitis. Samples were collected from Bulacan, Cagayan, Oriental Mindoro, Bohol and Cebu. Examinations were conducted by BFAR Central Fish Health Laboratory.</p>
11	<p>Acute Hepatopancreatic Necrosis Disease (AHPND) First Quarter: Origin of the disease or pathogen (history of the disease) - detected 5 farms Species affected: <i>P.vannamiei</i> (post-larvae, grow-out) Pathogen: Acute Hepatopancreatic Necrosis Disease Size of infected areas or names of infected areas: Negros Occidental, Cagayan, Occidental Mindoro, Oriental Mindoro and Cebu Samples sent to national or international laboratories for confirmation (indicate the name of the laboratories): Polymerase Chain Reaction (PCR) Test / BFAR Regional and SEAFDEC Fish Health Laboratories.</p> <p>Second Quarter: Origin of the disease or pathogen (history of the disease) - detected in 2 farms Species affected: <i>P. vannamiei</i> (post-larvae and grow-out) Pathogen: AHPND <i>Vibrio parahaemolyticus</i> Size of infected areas or names of infected areas: Oriental Mindoro and Leyte Samples sent to national or international laboratories for confirmation (indicate the name of laboratories): Polymerase Chain Reaction Test (PCR) / BFAR Regional Fish Health Laboratory.</p>
12	<p>Hepatopancreatic Microsporidiosis caused by <i>Enterocytozoon hepatopenaei</i> (HPM-EHP) First Quarter: Origin of the disease or pathogen (history of the disease) - detected 6 farms Species affected: <i>P.monodon</i> (grow-out) and <i>P.vannamiei</i> (fry, post-larvae, grow-out) Pathogen: Hepatopancreatic Microsporidiosis caused by <i>Enterocytozoon hepatopenaei</i> Size of infected areas or names of infected areas: Pampanga, Zambales, Quezon, Camarines Norte, Bohol and Cebu Samples sent to national or international laboratories for confirmation (indicate the name of the laboratories): Polymerase Chain Reaction (PCR) Test / BFAR Regional Fish Health Laboratories.</p> <p>Second Quarter: Origin of the disease or pathogen (history of the disease) - detected in 1 farm Species affected: <i>P. monodon</i> (post-larvae) Pathogen: Enterocytozoon hepatopenaei Size of infected areas or names of infected areas: Agusan del Norte Samples sent to national or international laboratories for confirmation (indicate the name of laboratories): Polymerase Chain Reaction Test (PCR) / BFAR Regional Fish Health Laboratory.</p>

2. New aquatic animal health regulations introduced within past six months (with effective date):